Global Sciences

Original Article

 \odot \odot

Investigation of *GJB2* and *SLC26A4* genes related to pendred syndrome genetic deafness patients

A B S T R A C T



Haider Majid Haider Al-Zaidi^{1,} ^(D), Fatemehsadat Mousavinasab², Nika Radseresht³, Ali Reza Mirzaei⁴, Yasaman Moradi³, Mohammad Mahmoudifar³

Deafness can occur due to damage to the ear, especially the inner

ear. In other cases, the cause is a heterogeneous genetic abnormality

and is caused by the changes that occur in the genes involved in the

hearing process. Mutations in GJB2 and SLC26A4 genes are one of

the most important causes of deafness in the world, which causes syndromic and non-syndromic hereditary hearing loss. The purpose of this study is to investigate *GJB2* and *SLC26A4* genes related to genetic syndromes of deafness and bioinformatic analysis at the genome and proteome level and to evaluate and compare the expression of these genes in different tissues of the human body. For this purpose, tools related to bioinformatics analysis such as UCSC and OMIM databases were used. One of the common genetic syndromes caused by mutations in these genes is pendred

syndrome. The clinical symptoms of this disease are weight gain,

constipation, dry skin, and hair, decreased energy, sleepiness, bulging belly, decreased body temperature, and slow growth. This disease does not currently have a specific treatment, so it is very important and fundamental to investigate the genetic factors affecting this disease. The results of this research showed that the transfer of potassium, sodium, and chlorine ions as well as the mutation in the *SLC26A4* gene, which is responsible for the synthesis of pendrin protein, is very effective in the occurrence of pendred syndrome. To diagnose pendred syndrome more accurately, molecular methods should be used in genetic tests. The results of comparing the expression profiles of these two genes showed that the difference in the expression of these two genes is very high and, in general, the expression of the *SLC26A4* gene in the body is very

low. Because people with hearing loss have other problems

including damage to other parts of the body such as the heart,

kidneys, or eyes. Knowing the genetic cause in these cases allows

the doctor to be aware of problems in other systems as well.

Article info Received: 02 Nov 2022 Revised: 08 Feb 2023 Accepted: 17 Mar 2023

Use your device to scan and read the article online



Keywords:

Bioinformatics Analysis, Thyroid Hormone, CX26 Mutant, Pendrin Protein, Connexin 26

1. Introduction

neurological diseases. More than 50% of deafness cases have a genetic origin. Pendred syndrome is a genetic and hereditary disorder

Deafness is one of the most common

¹Department of Otolaryngology and Skull Base Surgeon, College of Medicine, Ibn Sina University of Medical and Pharmaceutical Sciences, Baghdad, Iraq

²Institute for Biomedical Sciences, Georgia State University, Atlanta, GA, USA

³Department of Orthopaedic Dentistry, A. I. Evdokimov Moscow State University of Medicine and Dentistry, Moscow, Russia

⁴Department of Agronomy and Plant Breeding, Faculty of Agriculture and Natural Resources, University of Mohaghegh Ardabili, Ardabil, Iran

*Corresponding Author: Haider Majid Haider Al-Zaidi (hmh7895@gmail.com)

2023, 3(3): 163-171

that occurs as a result of defects in the function of the SLC26A4 gene. The pattern of this disease is autosomal recessive [1-4]. This syndrome is one of the syndromes associated with hearing loss and usually causes profound hearing loss, with an estimated prevalence of 1 in 14,500. This disease is caused by the thyroid hormone not working properly. It's most important symptom is hearing loss. In this disease, a person has hearing loss since birth, and the thyroid gland is enlarged and causing goiter. Goiter usually appears at the age of eight, but in some cases, it has been observed at birth [5]. As mentioned, the SLC26A4 gene encodes the pendrin protein. This protein is the cause of pendred syndrome. Pendrin protein is the transporter of sulfate inside and outside the cell, as well as the transporter of iodide and chloride in membrane cells [3, 6, 7]. Since potassium, sodium, and chlorine ions are very important for hearing, a defect in the SLC26A4 gene causes disruption in the transmission of sound waves to the brain and reduces the sense of hearing. Due to the lack of genetic research and gene therapy in this field, hearing aids and cochlear implants are currently used as effective treatment methods for children with this disorder (Figure 1). Other clinical symptoms of Pendred syndrome are weight gain, constipation, dry skin and hair, decreased energy, sleepiness, bulging belly, low body temperature, low growth, and mental retardation.

Research has shown that more than 150 gene loci are involved in deafness and more than 1200 different mutations in the human genome are involved in deafness [8]. According to the statistics of the World Health Organization, around 360 million people around the world have mild to severe deafness, and 1 out of every 1000 children born is deaf. Some environmental factors can cause hearing loss. Hearing loss in babies is due to "environmental" reasons, such as the mother's infection during pregnancy and complications after birth. In some cases, environmental genetic effects work together and cause hearing loss. Deafness has high heterogeneity, and in different clinical conditions, different genes play a role in the occurrence of this disease. GJB2 and SLC26A4 genes are common deafness genes and

account for 40% of genetic deafness patients [8, 9]. The purpose of this study is to investigate GJB2 and *SLC26A4* genes related to genetic syndromes of deafness and bioinformatic analysis at the genome and proteome level and to evaluate and compare the expression of these genes in different tissues of the human body.

2. Materials and methods

First, the sequences of GIB2 (NM_004004.6) and *SLC26A4* (NM_000441.2) genes were obtained from the NCBI database. The lengths of these proteins were 226 and 780 amino acids, respectively. The exact location of these genes was then determined using the UCSC database. The threedimensional structure of the proteins and drawing of a diagram of Ramachandran were determined using the MBC database, and the molecular weight and isoelectric point of the proteins were determined using the ProtScale database. Then, Cell comparison of GJB2 and *SLC26A4* genes and expression of these genes were examined by Human Protein Atlas OMIM database.



Fig. 1. Goiter associated with deafness and use of hearing aids and cochlear implants for children

3. Results

3.1. Analysis of genes related to deafness

The *GJB2* gene is located on chromosome number 13 and at position 13q12.11. It has 2 exons and 226 amino acids. This gene has a row of six consecutive G nucleotides at the position of nucleotides 30-35, which is prone to slippage in replication, which results in the premature termination codon in exon number 2. The most common type of mutation is the deletion of a nucleotide at the 30delG and 35delG position, which is the main cause of much hereditary and non-syndromic deafness (autosomal recessive in the DFNB1 locus and autosomal dominant in the DFNA3 locus). A single mutation in this gene, c.35delG, accounts for more than 60% of hereditary deafness in Northern European countries. Research has shown that the prevalence of the 35delG mutation is higher in the white population [10].

The *GJB2* gene provides direct intracellular communication and allows the passive release of molecules up to 1 kDa including nutrients, metabolites (glucose), ions (calcium and potassium), and secondary messengers such as (IP3, and cAMP)[<u>11</u>].

GJB2 gene encodes connexin 26 protein. Connexin 26 (CX26) is one of the main proteins involved in potassium (K+) homeostasis in the cochlea. CX26 is found in supporting cells, fibrocytes of the spiral ligament, and cells of the spiral limbus. If the CX26 protein is not enough, the potassium level in the inner ear will be too high, causing hearing damage [11]. It has suggested that deafness associated with CX26 mutations is caused not only by reduced potassium recirculation in the inner ear but also by abnormalities in the exchange of other metabolites through the cochlear cleft [12].

It has showed that CX26 and CX43 subunits are involved in the migration of neurons to the cerebral cortex, and the reduction of CX26 and CX43 at the points of contact between radial fibers and migrating neurons leads to neurological disorders in the cerebral cortex [13].

The *GJB6* gene encoding connexin 30 (CX30) is located near the *GJB2* gene. The proteins obtained from these two genes (*GJB2* and *GJB6*) are members of the gap junction protein family. This group includes 22 different genes that facilitate the transport of small molecules and ions between cells. Mutations in these two genes cause different degrees of deafness [<u>14</u>].

The *SLC26A4* gene is located on chromosome number 7 and at position 7q22.3. This gene is in the Solute carriers (SLC) group, which contains 433 different genes. It has 21 exons and 780 amino acids. This gene encodes the protein pendrin, which

is an anion transporter (chlorine and iodine ions). More than 300 mutations have been identified in this gene. A defect in this gene disrupts the function of the ear, thyroid, and kidney [15].

	SLC26A4		
Name	GJB2	SLC26A4	
Approved name	gap junction protein beta 2	solute carrier family 26 member 4	
ORGANISM	Homo sapiens (Human)	Homo sapiens (Human)	
Accession number nucleotide	NM_004004.6	NM_000441.2	
Accession number protein	NP_003995.2	NP_000432.1	
Gene ID	2706	5172	
Chromosome	13	7	
Cytogenetic location	13q12.11	7q22.3	
Chromosome	20187470-	107660828-	
location bp	20192938	107717809	
nucleotide length	2290 bp	4737 bp	
protein length	226aa	780aa	
Molecular weight (Da)	26215.07	85723.07	
Isoelectric point	9.11	6.04	
Total Exon	2	21	

3.2. Investigating the role of molecular biology and cell in the GJB2 gene

The *GIB2* gene plays a role in identical binding protein and causes cell communication, cell signaling. sound perception, and gap junction assembly in biological processes. Also, this gene plays an active role in the cellular components of the endoplasmic reticulum-Golgi intermediate compartment, membrane, plasma membrane, gap junction, connexin complex, and cell junction [<u>16-20</u>].

3.3. Investigating the role of molecular biology and cell in the *SLC26A4* gene

The molecular function of the *SLC26A4* gene is in the form of chloride channel activity, bicarbonate, chloride, iodide, sulfate, and oxalate transmembrane transporter activity, and secondary active sulfate transmembrane transporter activity [21-23]. Also, the *SLC26A4* gene plays a role in biological processes such as ion transport (sulfate, mineral anions, bicarbonate, iodide,

and oxalate), regulation of pH, especially intracellular pH, regulation of membrane potential, transmembrane transport (anion, chloride, and sulfate), regulation of protein localization and sensory perception of the sound [24-27]. Research has also shown that the *SLC26A4* gene is an integral part of the membrane and is actively present in the plasma membrane and extracellular exosome [28-30].

3.4. Three-dimensional structure

Molecular homologation modeling using the SWISS-MODEL server in Expasy resulted in a three-dimensional structure of GIB2 and *SLC26A4* proteins based on sample 1a02 with the highest similarity (Figure 2). The estimation of protein quality was determined according to the OMEANDisCo Global scale. OMEANDisCo global score [31] is the average per-residue QMEANDisCo score which has been found to correlate well with The Local Distance Difference Test (IDDT) score [32]. The provided error estimate is based on OMEANDisCo global scores estimated for a large set of models and represents the root mean squared difference (i.e. standard deviation) between QMEANDisCo global score and IDDT (the ground truth). As the reliability of the prediction depends on model size, the provided error estimate is calculated based on models of similar size to the input. The value of QMEANDisCo Global for the GIB2 gene is equal to 0.71±0.05 and for the SLC26A4 gene is equal to 0.45 ± 0.05 .

According to the QMEAN z-scores, it was found that there is a good match between the model structure and the experimental structures of the same size. Figures 3 and 4 show different QMEAN z-scores, which include QMEAN, C-beta interactions, and interactions between all atoms, solvation, and torsion, for *GJB2* and *SLC26A4* genes. Then Ramachandran diagram related to *GJB2* and *SLC26A4* proteins was determined to determine the energy level and stability in terms of two angles φ and ψ in proteins. Considering that in *GIB2* protein the percentage of amino acids in Ramachandran favoured was 94.65% and in SLC26A4 protein the percentage of amino acids of favoured 93.33%. Ramachandran was Therefore, the proposed model is suitable for three-dimensional structure for proteins (Figure 5).



Fig 2. A) Three-dimensional structure of *GJB2* protein. B) Three-dimensional structure of *SLC26A4* protein.

3.5. Gene expression analysis

Figures 6 and 7 show the analysis of *GIB2* and SLC26A4 gene expression in different body organs. The expression profile for the GIB2 gene showed that this gene has high expression in most organs and is not expressed only in the eye and connective and soft tissue. As shown in the figure, the *SLC26A4* gene is expressed only in endocrine tissues and is not expressed in other organs. According to the comparison of the expression profile of these two genes, we find that the difference in the expression of these two genes is very high and in general, the expression of the *SLC26A4* gene in the body is very low. The study of gene expression by microarray expression data also confirmed these findings.



QMEAN 2	Z-Sc	ores			
QMEAN				1	-4.80
Сβ				1	-4.66
All Atom				1	-1.99
solvation	1			1	-1.12
torsion			1 1	1	-3.88

Fig. 3. Plot showing the QMEAN value and Z-score for the *GJB2* gene



Fig. 4. Plot showing the QMEAN value and Z-score for the SLC26A4 gene



Fig. 5. The *GJB2* protein Ramachandran diagram (The right side), The *SLC26A4* protein Ramachandran diagram (left side)







The protein expression level of gene SLC26A4



Fig. 7. Expression profile of gene SLC26A4 in different organs of the body

4. Discussion

Manifestations of hearing are different; it is usually seen as moderate to profound sensor neural hearing loss. Hearing loss is usually diagnosed in the first two years of life and is often symmetrical. Considering the high prevalence of pendred syndrome as the second most common syndrome of deafness and also the frequency of consanguineous marriages, this disease, like any autosomal recessive disease, has a wide prevalence and the possibility of obtaining a molecular diagnosis, and genetic counseling seems essential for families [33]. So far, no significant relationship between the type and severity of goiter with the type of mutation in the SLC26A4 gene has been reported. Considering the widespread prevalence of goiter disease, its association with deafness alone does not indicate pendred syndrome unless a person with deafness and goiter shows a linkage to the DFNB4 locus. In these cases, it is necessary to perform other complementary thyroid tests such as thyroid function tests and perchlorate discharge tests in these people. Radiological changes of the inner ear and cochlea help to diagnose Pendred syndrome as a valuable diagnostic test, but the non-specificity of this test and the normalization of its findings do not alone determine the diagnosis of Pendred syndrome. Therefore, the totality of these findings indicates the necessity of conducting a molecular test to find the connection between the DFNB4 locus in non-syndromic deaf people [33].

It is worth mentioning that due to the different spectrum of identified mutations, it seems necessary to continue more extensive studies, including the study of the whole gene and studies on syndromic and non-syndromic hereditary deafness families. It is also necessary to evaluate the identified changes by DNA sequencing. This case is very effective for identifying other mutations of this locus in the deaf population. Understanding the genetic causes of deafness has important benefits. This knowledge not only allows doctors to inform families about their chances of having a hearing-impaired child, but it can also influence how people are treated for deafness [34].

5. Conclusion

The research conducted in this study along with the analysis of genes related to hearing in humans emphasizes the necessity of using bioinformatics in treatment. Analyzing data from genome sequencing, gene expression, and investigating mutations or gene variants that can affect the patient's response to a specific drug or change the prognosis of the disease is essential. The collection of these factors, along with the examination of databases and specialized software, has had a remarkable approach in the field of clinical trials and diagnosis and prevention of genetic diseases, which are expected to be used in the stages of gene therapy.

Conflict of Interests

All authors declare no conflict of interest.

Ethics approval and consent to participate

No human or animals were used in the present research.

Consent for publications

All authors read and approved the final manuscript for publication.

Informed Consent

The authors declare not used any patients in this research.

Availability of data and material

The data that support the findings of this study are available from the corresponding author upon reasonable request.

Authors' contributions

All authors had equal role in study design, work, statistical analysis and manuscript writing.

Funding

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

References

1. Hasnain MJU, Shoaib M, Qadri S, Afzal B, Anwar T, Abbas SH, Sarwar A, Talha Malik HM, Tariq Pervez M (2020) Computational analysis of functional single nucleotide polymorphisms associated with SLC26A4 gene. PLoS One 15 (1): e0225368. doi: https://doi.org/10.1371/journal.pone.022 5368

- 2. Kusano Y (2020) Pendred syndrome with hyperthyroidism. Journal of rural medicine
 : JRM 15 (4): 217-220. doi: https://doi.org/10.2185/jrm.2020-011
- 3. Lu YT, Wang L, Hou LL, Zheng PP, Xu Q, Deng DT (2022) SLC26A4 mutation in Pendred syndrome with hypokalemia: A case report. Medicine (Baltimore) 101 (35): e30253. doi: https://doi.org/10.1097/md.000000000 030253
- 4. Nonose RW, Lezirovitz K, de Mello Auricchio MTB, Batissoco AC, Yamamoto GL, Mingroni-Netto RC (2018) Mutation analysis of SLC26A4 (Pendrin) gene in a Brazilian sample of hearing-impaired subjects. BMC medical genetics 19 (1): 73. doi: <u>https://doi.org/10.1186/s12881-018-0585-x</u>
- 5. Dossena S, Rodighiero S, Vezzoli V, Nofziger C, Salvioni E, Boccazzi M, Grabmayer E, Bottà G, Meyer G, Fugazzola L (2009) Functional characterization of wild-type and mutated pendrin (SLC26A4), the anion transporter involved in Pendred syndrome. Journal of molecular endocrinology 43 (3): 93-103. doi: <u>https://doi.org/10.1677/JME-08-0175</u>
- 6. Matulevicius A, Bernardinelli E, Brownstein Z, Roesch S, Avraham KB, Dossena S (2022) Molecular Features of SLC26A4 Common Variant p.L117F. J Clin Med 11 (19). doi: https://doi.org/10.3390/jcm11195549
- Zhu K, Jin Y (2022) Case report: A case of SLC26A4 mutations causing pendred syndrome and non-cystic fibrosis bronchiectasis. Front Pediatr 10: 1077878. doi:

https://doi.org/10.3389/fped.2022.10778 78

8. Hilgert N, Smith RJ, Van Camp G (2009) Forty-six genes causing nonsyndromic hearing impairment: which ones should be analyzed in DNA diagnostics? Mutation Research/Reviews in Mutation Research 681 (2-3): 189-196. doi: https://doi.org/10.1016/j.mrrev.2008.08. 002 9. Abtahi SHR, Malekzadeh A, Soheilipour S, Salehi M, Taleban R, Rabieian R, Moafi M (2019) Evaluation of GJB2 and GJB6 mutations in patients afflicted with nonsyndromic hearing loss. International Journal of Pediatrics-Mashhad 7 (2): 9053-9060. doi: https://doi.org/10.22038/ijp.2018.34154.

3017 10. Rabionet R, Gasparini P, Estivill X (2000) Molecular genetics of hearing impairment due to mutations in gap junction genes encoding beta connexins. Human mutation 16 (3): 190-202. doi: https://doi.org/10.1002/1098-1004(200009)16:3<190::AID-HUMU2>3.0.CO;2-I

- 11. Valdebenito S, Barreto A, Eugenin EA (2018) The role of connexin and pannexin containing channels in the innate and acquired immune response. Biochimica et Biophysica Acta (BBA)-Biomembranes 1860 (1): 154-165. doi: https://doi.org/10.1016/j.bbamem.2017.0 5.015
- 12. Meşe G, Londin E, Mui R, Brink PR, White TW (2004) Altered gating properties of functional Cx26 mutants associated with recessive non-syndromic hearing loss. Human genetics 115: 191-199. doi: https://doi.org/10.1007/s00439-004-1142-6.
- 13. Elias LA, Wang DD, Kriegstein AR (2007) Gap junction adhesion is necessary for radial migration in the neocortex. Nature 448 (7156): 901-907. doi: https://doi.org/10.1038/nature06063
- 14. Wilch E, Azaiez H, Fisher RA, Elfenbein J, Murgia A, Birkenhäger R, Bolz H, Da Silva-Costa S, Del Castillo I, Haaf T (2010) A novel DFNB1 deletion allele supports the existence of a distant cis-regulatory region that controls GJB2 and GJB6 expression. Clinical genetics 78 (3): 267-274. doi: <u>https://doi.org/10.1111/j.1399-</u> 0004.2010.01387.x
- 15. Wilson G, Bryan J, Cranston K, Kitzes J, Nederbragt L, Teal TK (2017) Good enough practices in scientific computing. PLoS computational biology 13 (6): e1005510. doi:

https://doi.org/10.1371/journal.pcbi.1005 510

16. Appenzeller-Herzog C, Hauri H-P (2006) The ER-Golgi intermediate compartment (ERGIC): in search of its identity and function. Journal of cell science 119 (11): 2173-2183. doi: https://doi.org/10.1242/ics.03019

https://doi.org/10.1242/jcs.03019

- 17. Abascal F, Zardoya R (2013) Evolutionary analyses of gap junction protein families. Biochimica et Biophysica Acta (BBA)-Biomembranes 1828 (1): 4-14. doi: <u>https://doi.org/10.1016/j.bbamem.2012.0</u> 2.007
- 18. Kelsell DP, Dunlop J, Hodgins MB (2001) Human diseases: clues to cracking the connexin code? Trends in cell biology 11 (1): 2-6. doi: <u>https://doi.org/10.1016/S0962-</u> <u>8924(00)01866-3</u>
- 19. Shah MM, Landen CN (2014) Ovarian cancer stem cells: are they real and why are they important? Gynecologic oncology 132 (2): 483-489. doi: https://doi.org/10.1016/j.ygyno.2013.12.0 01
- 20. Höög JL, Lacomble S, Bouchet-Marquis C, Briggs L, Park K, Hoenger A, Gull K (2016) 3D architecture of the *Trypanosoma brucei* flagella connector, a mobile transmembrane junction. PLoS Neglected Tropical Diseases 10 (1): e0004312. doi: <u>https://doi.org/10.1371/journal.pntd.000</u> <u>4312</u>
- 21. Sindic A, Chang M-H, Mount DB, Romero MF (2007) Renal physiology of SLC26 anion exchangers. Current opinion in nephrology and hypertension 16 (5): 484-490. doi: <u>https://doi.org/10.1097/MNH.0b013e328</u> <u>2e7d7d0</u>
- 22. Kopp P, Pesce L, Solis-S JC (2008) Pendred syndrome and iodide transport in the thyroid. Trends in Endocrinology & Metabolism 19 (7): 260-268. doi: <u>https://doi.org/10.1016/j.tem.2008.07.00</u> 1
- 23. Alka K, Casey JR (2014) Bicarbonate transport in health and disease. IUBMB life 66 (9): 596-615. doi: https://doi.org/10.1002/iub.1315
- 24. Cherest H, Davidian J-C, Thomas D, Benes V, Ansorge W, Surdin-Kerjan Y (1997) Molecular characterization of two high affinity sulfate transporters in *Saccharomyces cerevisiae*. Genetics 145 (3): 627-635. doi:

https://doi.org/10.1093/genetics/145.3.6 27

25. Dror AA, Politi Y, Shahin H, Lenz DR, Dossena S, Nofziger C, Fuchs H, de Angelis MH, Paulmichl M, Weiner S (2010) Calcium Oxalate Stone Formation in the Inner Ear as a Result of an Slc26a4 Mutation. Journal of Biological Chemistry 285 (28): 21724-21735. doi: https://doi.org/10.1074/jbc.M110.120188

https://doi.org/10.1074/jbc.M110.120188

26. Bronckers AL, Guo J, Zandieh-Doulabi B, Bervoets TJ, Lyaruu DM, Li X, Wangemann P, DenBesten P (2011) Developmental expression of solute carrier family 26A member 4 (SLC26A4/pendrin) during amelogenesis in developing rodent teeth. European journal of oral sciences 119: 185-192. doi: https://doi.org/10.1111/j.1600-

0722.2011.00901.x

- 27. Ishimori S, Kaito H, Matsunoshita N, Otsubo H, Hashimoto F, Ninchoji T, Nozu K, Morisada N, Iijima K (2013) SLC26A3 gene analysis in patients with Bartter and Gitelman syndromes and the clinical characteristics of patients with unidentified mutations. Kobe J Med Sci 59 E36-E43. doi: (2): https://doi.org/10.1016/j.mam.2012.07.0 09
- 28. Witwer KW, Buzás EI, Bemis LT, Bora A, Lässer C, Lötvall J, Nolte-'t Hoen EN, Piper Sivaraman (2013)MG. S, Skog Ι Standardization of sample collection, isolation and analysis methods in extracellular vesicle research. Journal of extracellular vesicles 2 (1): 20360. doi: https://doi.org/10.3402/jev.v2i0.20360
- 29. Février B, Raposo G (2004) Exosomes: endosomal-derived vesicles shipping extracellular messages. Current opinion in cell biology 16 (4): 415-421. doi: https://doi.org/10.1016/j.ceb.2004.06.003
- 30. Teng Y, Ren Y, Hu X, Mu J, Samykutty A, Zhuang X, Deng Z, Kumar A, Zhang L, Merchant ML (2017) MVP-mediated exosomal sorting of miR-193a promotes colon cancer progression. Nature communications 8 (1): 14448. doi: https://doi.org/10.1038/ncomms14448
- 31. Studer G, Rempfer C, Waterhouse AM, Gumienny R, Haas J, Schwede T (2020) QMEANDisCo—distance constraints applied on model quality estimation.

Bioinformatics 36 (6): 1765-1771. doi: https://doi.org/10.1093/bioinformatics/b tz828

- 32. Mariani V, Biasini M, Barbato A, Schwede T (2013) IDDT: a local superposition-free score for comparing protein structures and models using distance difference tests. Bioinformatics 29 (21): 2722-2728. doi: https://doi.org/10.1093/bioinformatics/b tt473
- 33. Honda K, Griffith AJ (2022) Genetic architecture and phenotypic landscape of *SLC26A4*-related hearing loss. Human Genetics 141 (3-4): 455-464. doi: <u>https://doi.org/10.1007/s00439-021-</u> <u>02311-1</u>
- 34. Chouchen J, Mahfood M, Alobathani M, Mohamed WKE, Tlili A (2021) Clinical heterogeneity of the SLC26A4 gene in UAE patients with hearing loss and bioinformatics investigation of **DFNB4/Pendred** syndrome missense mutations. International Journal of Pediatric Otorhinolaryngology 140: 110467. doi: https://doi.org/10.1016/j.jporl.2020.110 467

Copyright © 2023 by the author(s). This is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/licenses/by/4.0/)

How to Cite This Article:

Al-Zaidi HMH, Mousavinasab F, Radseresht N, Mirzaei AR, Moradi Y, Mahmoudifar M (2023) Investigation of GJB2 and SLC26A4 genes related to pendred syndrome genetic deafness patients. Cellular, Molecular and Biomedical Reports 3 (3): 163-171. doi: 10.55705/cmbr.2023.379262.1093

Download citation:

RIS; EndNote; Mendeley; BibTeX; APA; MLA; HARVARD; VANCOUVER