Review Article

Comprehensive analysis of microRNA (miRNA) in cancer cells



Naghmana Kanwal¹, Othman Rashid Al-Samarrai², Haider Majid Haider Al-Zaidi³, Ali Reza Mirzaei⁴,*[®], Mohammad Javad Heidari⁵

MicroRNAs (miRNA) are a group of small non-coding RNAs that

regulate gene expression at the RNA level. MicroRNAs have positive

regulatory effects on protein translation processes and often induce

their performance by binding to the 3'-UTR mRNA region. Also, microRNAs are involved in various cellular processes, including development, cell division, cell signaling, and cell growth, and generally play an effective role in the cell cycle and control of

physiological processes and cell pathology. Several studies confirm

that microRNAs play an important role in the initiation and progression of cancer, and many of them act as oncogenes and tumor suppressors. On the other hand, microRNAs are important stimulating factors that can act as biomarkers in the diagnosis and prognosis of various types of cancer, and in many cases, the occurrence of mutations in microRNAs and open-reading templates can lead to cancer. MicroRNAs also play an effective role in regulating gene expression. Biological studies have shown that

about 30% of all genes and the majority of genetic pathways are

regulated by microRNAs. In general, microRNAs and their target

molecules are potential biological goals for primary screening,

targeted treatment, and pharmaceutical resistance, and identifying

them provides a clear prospect for a better understanding of the

ABSTRACT

Citobal Sciences Article info Received: 05 Apr 2022 Revised: 27 Aug 2022 Accepted: 30 Oct 2022

Use your device to scan and read the article online



Keywords: MiRNA-21, Tumor Inhibitor, DNA Methyltransferase Oncogene, Reduction of Apoptosis, Intragenic miRNA

1. Introduction

MicroRNAs are a group of non-coding, intragenic, and single-stranded RNAs with a length of 18-25 nucleotides, which are complementary to mRNAs of protein-coding genes and can prevent the expression of related genes and proteins. MicroRNA biogenesis takes place in the nucleus and cytoplasm. Considering the role of MicroRNAs of in the process proliferation and differentiation, it is expected that the disruption of the expression of MicroRNAs is

related to cancer. Oncomirs are a group of MicroRNAs that can act as oncogenes or tumor suppressors by reducing the expression of cancer-related target genes [1-3].

More than 50% of microRNA genes are present in genomic regions related to cancer or fragile areas, which undergo the process of deletion or duplication, which indicates their role in malignant transformations [4-6]. MicroRNAs control gene expression after

pathways leading to cancer.

¹Department of Health Care Biotechnology, Atta-ur-Rahman School of Applied Biosciences, National University of Sciences and Technology, H-12, Islamabad, Pakistan

²Applied Chemistry Department, College of Applied Science, University of Samarra, Samarra, Iraq

³Department of Surgery, College of Medicine, Ibn Sina University of Medical and Pharmaceutical Sciences, Baghdad, Iraq

⁴Department of Agronomy and Plant Breeding, Faculty of Agriculture and Natural Resources, University of Mohaghegh Ardabili, Ardabil, Iran

⁵Faculty of Pharmacy, Cyprus International University, Lefkosa, Turkish Republic of Northern Cyprus, Lefkosa, Cyprus *Corresponding Author: Ali Reza Mirzaei (<u>amirzaei25@amail.com</u>)

transcription by binding to the region of 3'-UTR mRNA by inhibiting the translation of mRNAs or inducing their degradation. The primary transcripts of microRNAs are converted into mature microRNAs during 2 processing steps [4, 5]. MicroRNAs were first discovered in nematodes in 1993 by Li et al. and subsequently identified in chronic lymphocytic leukemia [7].

Subsequent studies showed that the region contains 2 genes encoding miRNA-16-1 and miRNA-15a, which are transcribed polycistronically [8].

In 2004, microchip analysis was used to characterize miRNAs in humans and mice. The microchips contained 245 different miRNAs that were identified by Northern blot and RT-PCR methods. MicroRNAs participate in many reactions and play an active role in various cellular processes, including cell signaling and cell growth and division. MicroRNAs also participate in physiological processes such as apoptosis, insulin secretion, blood supply, and tissue differentiation. Clinical studies have shown that MicroRNAs play an effective role in defending the body's immune system, especially against viral diseases [<u>9</u>]. MicroRNAs are transcribed bv RNA polymerase II in the nucleus. Then this 2stranded precursor is converted by Drosha, which is a type of endonuclease III, into a stem-loop structure with a length of 60-75 nucleotides. Preformed miRNA is transferred to the cytoplasm by Exportin 5 protein and processed by cutting enzymes. Then, by binding to the AGO protein, it turns into the RISC silencing complex. The binding site of mature miRNA is the 3'-UTR mRNA region of the target gene. In general, miRNAs inhibit protein synthesis. Target gene regulation by miRNAs depends on the degree of complementarity of miRNAs with the 3'-UTR mRNA region. MiRNAs with high degrees of similarity break the target gene, while lower degrees of similarity can cause gene silencing or decrease the expression of the target gene (figure 1)[10-12].

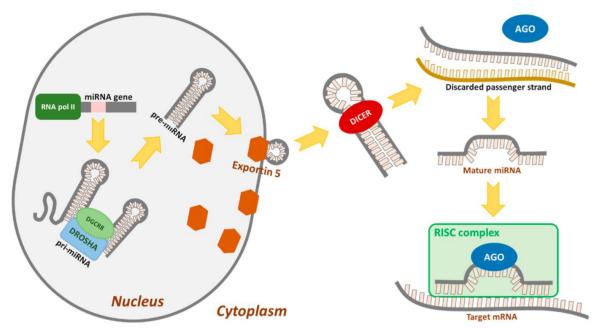


Fig. 1. Schematic illustration of miRNA synthesis process [<u>13</u>]. This figure is under CC BY license (<u>https://creativecommons.org/licenses/by/4.0/</u>).

2. Investigating the expression of MicroRNAs in cancer cells

MicroRNAs show different behavior in cancer cells. A group of MicroRNAs known as oncomirs is overexpressed in cancer cells.

This action causes tumor inhibitors to decrease in cancer cells. MiRNA-21 is overexpressed in most cancers, which decreases tumor suppressors such as PDCD4, PTEN, and TPM1 [14-16]. Another group of

microRNAs in cancer cells has a decrease in expression. Since the targets of these microRNAs are proto-oncogenes, the reduction in the expression of these microRNAs leads to the increase of protooncogenes in the cell. Let7 is reduced in most cancer cells, and RAS proto-oncogene is one of the targets of this microRNA [17, 18]. Another

group of MicroRNAs targets DNA methyltransferase. Decreased expression of these MicroRNAs leads to reduced expression and lowers tumor suppressors in cancer cells. In general, MicroRNAs that have decreased expression are called tsmiRNAs. Table 1 shows the types of MicroRNAs that are disrupted in cancer cells [19, 20].

Table 1. Types of MicroRNAs that are disrupted in cancer cells

MicroRNAs	target gene	consequences	Expression
MiRNA-21	Tumor inhibitor	Reduction of PTEN, PDCD4, reduction of apoptosis, and increase of cell growth	Increased expression
MiRNA-10b	Tumor inhibitor	Reduction of HOXD10, increase in invasion, and cell migration.	Increased expression
MiRNA-373	Tumor inhibitor	Reduction of CD44, reduction of cell attachment to the substrate	Increased expression
Let-7	DNA methyltransferase Oncogene	Increasing RAS, increasing cell proliferation	Reduced expression
MiRNA-206	DNA methyltransferase Oncogene	Increasing ER-alpha, increasing cell proliferation	Reduced expression
MiRNA- 450a	DNA methyltransferase Oncogene	Increasing DNMT, Epigenetic silencing of tumor suppressor genes	Reduced expression

3. DNA methylation

DNA methylation occurs normally in all vertebrate cells. CpG Island in the gene promoter plays an important role in the methylation and transcription of miRNAs. In this process, carbon number 5 of the cytosine ring in CpG dinucleotides plays an essential role. Methylation is done by DNMT (DNA methyltransferase) and using (S-adenosylmethionine) SAM as a methylating group. In normal cells, miRNAs associated with CpG islands are usually unmethylated. These miRNAs lead to histone configuration in euchromatin and provide access to TFs, H3K4, Co-Act, HATs, and HMT factors and form the transcription complex that includes RNA polymerase II, TAF (TBP-associated factor), TBP(TATA-binding protein) and finally transcription of primary and mature miRNAs is done. In tumor-bearing cells, promoter CpG with intergenic Island associated and miRNAs intragenic is abnormally hypermethylated. In this process, DNMT using HMTs, HDAC, and MBD leads to compact chromatin structure and finally, access to transcription factors is blocked and miRNAs are turned off [21-25].

4. Identification of target molecules in microRNAs

One of the most important topics related to microRNAs is the identification of their target molecules. MicroRNAs usually attach to the 3'-UTR end of mRNA. For this reason, the isolation of full-length genomics has been expanded in the last few decades [26]. In the complete isolation of the gene, the Rapid Amplification of cDNA Ends (RACE) technique is used, in this technique, in addition to the central CDS region, the 3'-UTR and 5'-UTR regions are also isolated [27]. The presence of complementary base pairs is necessary for the interaction between microRNAs and the sequence of the target molecule. In these interactions. usually 2-9 nucleotides at the 5' end of microRNAs, called the seed region, are paired with 6-7 nucleotides at the 3'-UTR end [<u>26</u>].

Other base pairs of microRNAs show a limited connection with the 3'-UTR sequence, which makes microRNAs able to bind to several positions in the 3'-UTR. Many software and computer algorithms have been used to detect the target location of microRNAs from the seed sequence. Bioinformatics studies have shown that more than a third of the human genome is regulated

by microRNAs, and about 1000 microRNAs have been estimated for one percent of the human genome [28-33].

5. MicroRNAs and cancer

Cancer is the result of the departure of cells from the normal path. Avoiding cell death, unlimited proliferative potential. tissue invasion. and metastasis lead to the development of malignant cancers. The effect microRNAs in the development of of programmed cell death, differentiation, and proliferation has been proven. The structure and function of microRNAs show that some microRNAs are abnormally expressed in cancer. There is also a significant relationship between the performance of tumors in different stages of cancer and the expression of microRNAs. However, in different cancers, the expression level of microRNAs is different, which can be caused by the difference between the origin of cancer cells and the surrounding stromal tissue (Table 2) [<u>34</u>].

In general, microRNAs can act as regulators of epigenetic factors. This is even though epigenetic factors also cause the inactivation of microRNAs through excessive methylation or histone changes. Gene deletions, exogenous silencers, and nonexpression of transcription factors also decrease the expression of tumor-inhibiting microRNAs [35]. In general, microRNAs lead to the reduction of tumor genesis and cancer by targeting proteogenic mRNAs and silencing them, reducing the expression of tumor genes, suppressor regulating cell differentiation, and programming cell death [34].

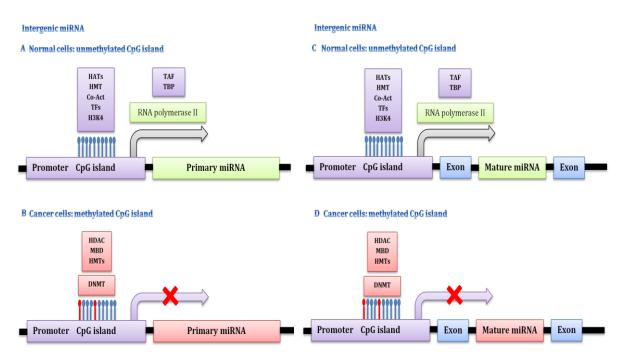


Fig. 2. A schematic model of DNA methylation-dependent miRNAs in normal & cancer cells [21, 22]. A. Normal cells: unmethylated CpG island (Intergenic miRNA) B. Cancer cells: methylated CpG island (Intergenic miRNA) C. Normal cells: unmethylated CpG island (Intragenic miRNA) D. Cancer cells: methylated CpG island (Intragenic miRNA). TF: Transcription factor; Pol II: RNA polymerase II; HDAC: Histone deacetylase; MBD: Methyl-CpG-binding domain protein; TAF: TBP-associated factor; TBP: TATA-binding protein; Co-Act: Co-activator; HMT: Histone methyltransferase; DNMT: DNA methyltransferase; HAT: Histone acetyltransferase.

Table 2. Changes in microRNA expression in different human cancers [34]

Types of cancer	Increased expression	Reduced expression
Breast	miR-21, miR-22, miR-23, miR-29b-2, miR-96, miR-155, miR191, miR-181, miR-182, miR-27a, miR-210	miR-205, miR-143, miR-145, miR10b, miR- 125a/b, miR-155, miR17-5p, miR-27b, miR-9-3, miR-31, miR-34 family, let-7
Ovarian	miR-200 a/b/c, miR-141, miR-18a, miR-93, miR-429	miR-199a, miR-140, miR-145, miR-125a,b, let7
Colorectal	miR-18, miR-224, miR-10a, miR-17-92 cluster, miR-21, miR-24-1, miR29b-2, miR-31, miR-96, miR-135b, miR- 183	miR-143, miR-145, let-7, miR30 -3p, miR-124a, miR-129, miR133 b, miR328
Lung	miR-21, miR-155, miR-191, miR 205, miR-210 miR-17- 92 cluster	let-7, miR-34 family, miR-143, miR-145, miR-124a
Glioblastoma	miR-221, miR-222, miR-21	miR-181a, miR-181b, miR-181c, miR-125a, miR- 125b
Esophageal cancer	miR-194, miR-192, miR-200c, miR-21	miR-203, miR-205
Stomach-intestinal	miR-106b-25	miR-15b, miR-16
Pancreas	miR-221, miR-376a, miR301, miR-21, miR-24-2, miR- 100, miR-103, miR107,miR125b-1, miR-155, miR-181, miR-106, miR-363, miR-301, miR-212, miR-34a376 miR-146h miR-221 miR-222 miR-181h miR-155 miR-	miR-375, let-7, miR-200, miR200b
thyroid	miR-146b, miR-221, miR-222, miR-181b, miR-155, miR- 197, miR-224, miR-346	miR-30d, miR-125b, miR-26a,miR-30a-5p
prostate	let-7d, miR-195, miR-203, miR-21,miR-181, miR-106, miR-363, miR-221	miR-128a, miR-101, miR-125a/b, miR-15a, miR-16-1, miR-143, miR-145, miR-23a/b, miR- 200, miR-330, miR-331
Bladder	miR-17, miR-23a,b, miR-26b, miR-103-1, miR-185, miR- 203, miR-205, miR-221, miR-223	miR-29c, miR-26a, miR-30c, miR-30e-5p, miR-145, miR-30a-3p, miR-133a/b, miR-195, miR125b, miR-199a

6. Conclusion

Approximately, 2694 mature human microRNAs have been identified, which have a significant impact on many human diseases, including neurological disorders. cardiovascular problems, diabetes, and cancer [36-41]. Today, microRNAs are widely used in cancer treatment and radiation therapy (RT). The results have shown that the high expression of miR-20b-5p in tumor cells makes these cells more sensitive to radiation therapy, which has led to an increase in the efficiency of this treatment method [42]. Xiang et al. showed that miR-139-5p in bone marrow mesenchymal stem cells (BMSCs) delays tumorigenesis and metastasis of bladder cancer cells in vivo by inactivating the p21 factor [43]. Also, microRNAs have an important regulatory role in diseases and are used as promising biomarkers for cancer diagnosis and prognosis. For example, miRNA-21 and miRNA-155 in serum samples can identify people with breast cancer from normal people with 100% accuracy [44]. Some microRNAs are disrupted in SARS infection. In general, microRNAs can interact with other molecules such as viruses and bacteria, and act as mediators for viral infections. Analysis of microRNA expression profiles and RNA-seq in COVID-19 patients showed that MIR302C-5p microRNA 302c-5p [Homo sapiens (human)] (hsa-miR-302c-5p)

and MIR16-5p microRNA 16-5p [Homo sapiens (human)] (hsa-miR-16-5p) are related to the regulation of pro-inflammatory cytokines, maturation, and differentiation of immune cells [45]. MicroRNAs can specifically bind to the target gene and repress it. The therapeutic performance of microRNAs in human trials largely depends on their incorporation into the target site and appropriate concentrations. Nanocarrier systems have a high potential to overcome challenges. Nanocarrier systems existing provide high stability during systemic circulation, efficient loading, and protection of microRNAs against nucleases, endosomal escape, and specific cell targeting. It is expected that microRNA-based nanotherapies will be used in cancer treatment in the next 5 years [<u>46</u>].

In general, microRNAs are important functional molecules that are included in the group of non-coding RNAs. The applications of microRNAs in the cell are so many that it sometimes makes it difficult to recognize their function. It is possible that the expression of microRNAs can be regulated through the synthesis of pre-microRNAs or antisense oligonucleotides. Also, microRNAs participate in many pathological and biological cellular processes, and the occurrence of mutations in them can lead to cancer. Several studies confirm that microRNAs play an important role in the initiation and progression of cancer, and depending on the type of microRNAs they control, they can inhibit tumors or oncogenes, which show a promising perspective for cancer treatment [47]. In general, these factors have caused scientists to use microRNAs as an important medicinal agent in the treatment of many diseases.

Conflict of Interest

All authors state that they are free of any conflicts of interest related to this paper.

Author's contributions

All authors confirm that they have read and approved this manuscript.

Consent for publications

All authors have read and approved the final manuscript for publication.

Availability of data and material

The authors have embedded all data in the manuscript.

Ethics approval and consent to participate

The authors did not use human or animals in the research.

Funding

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

References

1. López-Urrutia E, Bustamante Montes LP, Ladrón de Guevara Cervantes D, Pérez-Plasencia C, Campos-Parra AD (2019) Crosstalk between long non-coding RNAs, micro-RNAs and mRNAs: deciphering molecular mechanisms of master regulators in cancer. Frontiers in oncology 9): 669. doi:https://doi.org/10.3389/fonc.2019.00

<u>669/full</u>
2. Amodio F, Caiazza M, Fimiani F, Calabrò P, Limongelli G (2021) MicroRNAs: From Junk RNA to Life Regulators and Their Role in Cardiovascular Disease. Cardiogenetics 11 (4): 230-254. doi:https://doi.org/10.3390/cardiogenetic s11040023

- 3. Ren B, Guan M-X, Zhou T, Cai X, Shan G (2022) Emerging functions of mitochondria-encoded noncoding RNAs. Trends in Genetics). doi:<u>https://doi.org/10.1016/j.tig.2022.08.</u> 004
- Melo SA, Esteller M (2011) Dysregulation of microRNAs in cancer: Playing with fire. FEBS Letters 585 (13): 2087-2099. doi:<u>https://doi.org/10.1016/j.febslet.2010</u>.08.009
- 5. Lundstrom K (2011) Micro-RNA in disease and gene therapy. Current Drug Discovery Technologies 8 (2): 76-86. doi:<u>https://doi.org/10.2174/1570163117</u> 95563857
- 6. Wang W, Hao L-P, Song H, Chu X-Y, Wang R (2022) The potential roles of exosomal non-coding RNAs in hepatocellular carcinoma. Frontiers in Oncology 12): Article 790916. doi:<u>https://doi.org/10.3389/fonc.2022.79</u>0916
- 7. Mirnezami AHF, Pickard K, Zhang L, Primrose JN, Packham G (2009) MicroRNAs: Key players in carcinogenesis and novel therapeutic targets. European Journal of Surgical Oncology (EJSO) 35 (4): 339-347.

doi:https://doi.org/10.1016/j.ejso.2008.06 .006

- 8. Benetatos L, Vartholomatos G (2012) Deregulated microRNAs in multiple myeloma. Cancer 118 (4): 878-887. doi:https://doi.org/10.1002/cncr.26297
- Liu C-G, Calin GA, Meloon B, Gamliel N, Sevignani C, Ferracin M, Dumitru CD, Shimizu M, Zupo S, Dono M (2004) An oligonucleotide microchip for genomewide microRNA profiling in human and mouse tissues. Proceedings of the National Academy of Sciences 101 (26): 9740-9744. doi:https://doi.org/10.1073/pnas.040329 3101
- Hashemipour M, Boroumand H, Mollazadeh S, Tajiknia V, Nourollahzadeh Z, Rohani Borj M, Pourghadamyari H, Rahimian N, Hamblin MR, Mirzaei H (2021) Exosomal microRNAs and exosomal long non-coding RNAs in gynecologic cancers. Gynecologic Oncology 161 (1): 314-327.

doi:https://doi.org/10.1016/j.ygyno.2021. 02.004

- Rahaman M, Mukherjee M, Bhattacharya S, Mukherjee B, Shukla PC, Dolai TK, Chakravorty N (2022) Exploring the crosstalk between long non-coding RNAs and microRNAs to unravel potential prognostic and therapeutic biomarkers in β-thalassemia. Molecular Biology Reports): 1-12. doi:<u>https://doi.org/10.1007/s11033-022-07629-1</u>
- Ramalingam H, Yheskel M, Patel V (2020) Modulation of polycystic kidney disease by non-coding RNAs. Cellular Signalling 71): 109548. doi:<u>https://doi.org/10.1016/j.cellsig.2020.</u>

<u>109548</u>

- 13. Gajek A, Gralewska P, Marczak A, Rogalska A (2021) Current implications of microRNAs in genome stability and stress responses of ovarian cancer. Cancers 13 (11): 2690. doi:<u>https://doi.org/10.3390/cancers1311 2690</u>
- 14. Ghasabi M, Mansoori B, Mohammadi A, Duijf PH, Shomali N, Shirafkan N, Mokhtarzadeh A, Baradaran B (2019) MicroRNAs in cancer drug resistance: Basic evidence and clinical applications. Journal of cellular physiology 234 (3): 2152-2168. doi:<u>https://doi.org/10.1002/jcp.26810</u>
- 15. Omar HA, El-Serafi AT, Hersi F, Arafa ESA, Zaher DM, Madkour M, Arab HH, Tolba MF (2019) Immunomodulatory MicroRNAs in cancer: targeting immune checkpoints and the tumor microenvironment. The FEBS Journal 286 (18): 3540-3557. doi:<u>https://doi.org/10.1111/febs.15000</u>
- 16. Fazeli-Nasab B, Sayyed RZ, Sobhanizadeh A (2021) In Silico Molecular Docking Analysis of α -Pinene: An Antioxidant and Anticancer Drug Obtained from *Myrtus communis*. Int J Cancer Manag 14 (2): e89116.

doi:<u>https://doi.org/10.5812/ijcm.89116</u>

17. Tokumaru Y, Asaoka M, Oshi M, Katsuta E, Yan L, Narayanan S, Sugito N, Matsuhashi N, Futamura M, Akao Y (2020) High expression of microRNA-143 is associated with favorable tumor immune microenvironment and better survival in estrogen receptor positive breast cancer. International journal of molecular sciences 21 (9): 3213. doi:<u>https://doi.org/10.3390/ijms2109321</u> <u>3</u>

- 18. Xing Y, Ruan G, Ni H, Qin H, Chen S, Gu X, Shang J, Zhou Y, Tao X, Zheng L (2021) Tumor immune microenvironment and its related miRNAs in tumor progression. Frontiers in Immunology 12): 624725. doi:<u>https://doi.org/10.3389/fimmu.2021. 624725/full</u>
- 19. Lai X, Eberhardt M, Schmitz U, Vera J (2019) Systems biology-based investigation of cooperating microRNAs as monotherapy or adjuvant therapy in cancer. Nucleic acids research 47 (15): 7753-7766.

doi:<u>https://doi.org/10.1093/nar/gkz638</u>

20. Xie X, Pan J, Han X, Chen W (2019) Downregulation of microRNA-532-5p promotes the proliferation and invasion of bladder cancer cells through promotion of HMGB3/Wnt/β-catenin signaling. Chemico-Biological Interactions 300): 73-81.

doi:<u>https://doi.org/10.1016/j.cbi.2019.01.</u> 015

- 21. Wong KY, Yu L, Chim CS (2011) DNA methylation of tumor suppressor miRNA genes: a lesson from the *miR-34* family. Epigenomics 3 (1): 83-92. doi:https://doi.org/10.2217/epi.10.74
- 22. Wang LQ, Chim CS (2015) DNA methylation of tumor-suppressor miRNA genes in chronic lymphocytic leukemia. Epigenomics 7 (3): 461-473. doi:https://doi.org/10.2217/epi.15.6
- 23. Lewis B, Burge C (2005) Bartel DPJc. Conserved seed pairing, often flanked by adenosines, indicates that thousands of human genes are microRNA targets 120 (1): 15-20. doi:<u>https://doi.org/10.1016/j.cell.2004.12.</u> 035
- 24. Li Z, Wong KY, Calin GA, Chng W-J, Chan GC-f, Chim CS (2019) Epigenetic silencing of miR-340-5p in multiple myeloma: mechanisms and prognostic impact. Clinical epigenetics 11 (1): 1-13. doi:https://doi.org/10.1186/s13148-019-0669-2
- 25. Saito Y, Liang G, Egger G, Friedman JM, Chuang JC, Coetzee GA, Jones PA (2006) Specific activation of microRNA-127 with downregulation of the proto-oncogene *BCL6* by chromatin-modifying drugs in

human cancer cells. Cancer Cell 9 (6): 435-443.

doi:https://doi.org/10.1016/j.ccr.2006.04. 020

- 26. Kanellopoulou C, Monticelli S (2008) A role for microRNAs in the development of the immune system and in the pathogenesis of cancer. Seminars in Cancer Biology 18 (2): 79-88. doi:<u>https://doi.org/10.1016/j.semcancer.2008.01.002</u>
- 27. Lan X, Ruminy P, Bohers E, Rainville V, Viennot M, Viailly P-J, Etancelin P, Tilly H, Mihailescu S, Bouclet F, Leprêtre S, Jardin F (2022) 5' Rapid amplification of cDNA ends (5'RACE): A simpler method to analyze immunoglobulin genes and discover the value of the light chain in chronic lymphocytic leukemia. Leukemia Research 123): 106952. doi:<u>https://doi.org/10.1016/j.leukres.202</u> 2.106952
- 28. Babashah S, Soleimani M (2011) The oncogenic and tumour suppressive roles of microRNAs in cancer and apoptosis. European Journal of Cancer 47 (8): 1127-1137.
 doi:<u>https://doi.org/10.1016/j.ejca.2011.02</u>.008
- 29. Bűssing I, Slack FJ (2008) a Großhans, Helge. let-7 microRNAs in development, stem cells and cancer. Trends in molecular medicine 14 (9): 400-409. doi:<u>https://doi.org/10.1016/j.molmed.200</u> 8.07.001
- 30. Cho WCS (2010) MicroRNAs in cancer from research to therapy. Biochimica et Biophysica Acta (BBA) - Reviews on Cancer 1805 (2): 209-217. doi:<u>https://doi.org/10.1016/j.bbcan.2009. 11.003</u>
- 31. Cho WCS (2010) MicroRNAs: Potential biomarkers for cancer diagnosis, prognosis and targets for therapy. The International Journal of Biochemistry & Cell Biology 42 (8): 1273-1281. doi:https://doi.org/10.1016/j.biocel.2009. 12.014
- 32. Giovannetti E, Erozenci A, Smit J, Danesi R, Peters GJ (2012) Molecular mechanisms underlying the role of microRNAs (miRNAs) in anticancer drug resistance and implications for clinical practice. Critical Reviews in Oncology/Hematology

81 (2): 103-122. doi:<u>https://doi.org/10.1016/j.critrevonc.2</u> 011.03.010

- 33. Hanahan D, Weinberg Robert A (2011) Hallmarks of Cancer: The Next Generation. Cell 144 (5): 646-674. doi:https://doi.org/10.1016/j.cell.2011.02. 013
- 34. Schaefer A, Jung M, Kristiansen G, Lein M, Schrader M, Miller K, Stephan C, Jung K (2010) MicroRNAs and cancer: Current state and future perspectives in urologic oncology. Urologic Oncology: Seminars and Original Investigations 28 (1): 4-13. doi:<u>https://doi.org/10.1016/j.urolonc.200</u> 8.10.021
- 35. Ruan K, Fang X, Ouyang G (2009) MicroRNAs: Novel regulators in the hallmarks of human cancer. Cancer Letters 285 (2): 116-126. doi:<u>https://doi.org/10.1016/j.canlet.2009.</u> 04.031
- 36. Nguyen D-D, Chang S (2017) Development of novel therapeutic agents by inhibition of oncogenic microRNAs. International journal of molecular sciences 19 (1): 65. doi:<u>https://doi.org/10.3390/ijms1901006</u> 5
- 37. Wu K, He J, Pu W, Peng Y (2018) The Role of Exportin-5 in MicroRNA Biogenesis and Cancer. Genomics, Proteomics & Bioinformatics 16 (2): 120-126. doi:<u>https://doi.org/10.1016/j.gpb.2017.09</u> .004
- 38. Miller BH, Wahlestedt C (2010) MicroRNA dysregulation in psychiatric disease. Brain Research 1338): 89-99. doi:<u>https://doi.org/10.1016/j.brainres.20 10.03.035</u>
- 39. Friedman RC, Farh KK-H, Burge CB, Bartel DP (2009) Most mammalian mRNAs are conserved targets of microRNAs. Genome research 19 (1): 92-105. doi:<u>https://doi.org/10.1101/gr.082701.10</u> 8
- 40. Guay C, Roggli E, Nesca V, Jacovetti C, Regazzi R (2011) Diabetes mellitus, a microRNA-related disease? Translational Research 157 (4): 253-264. doi:<u>https://doi.org/10.1016/j.trsl.2011.01.</u> 009
- 41. Calin GA, Croce CM (2006) MicroRNA signatures in human cancers. Nature

reviews cancer 6 (11): 857-866. doi:<u>https://doi.org/10.1038/nrc1997</u>

- 42. Jiang K, Zou H (2022) microRNA-20b-5p overexpression combing Pembrolizumab potentiates cancer cells to radiation therapy via repressing programmed death-ligand 1. Bioengineered 13 (1): 917-929. doi:<u>https://doi.org/10.1080/21655979.20 21.2014617</u>
- 43. Xiang Y, Lv D, Song T, Niu C, Wang Y (2022) Tumor suppressive role of microRNA-139-5p in bone marrow mesenchymal stem cells-derived extracellular vesicles in bladder cancer through regulation of the KIF3A/p21 axis. Cell Death & Disease 13 (7): 1-12. doi:https://doi.org/10.1038/s41419-022-04936-0
- 44. Weng S, Lin D, Lai S, Tao H, Chen T, Peng M, Qiu S, Feng S (2022) Highly sensitive and reliable detection of microRNA for clinically disease surveillance using SERS biosensor integrated with catalytic hairpin assembly amplification technology. Biosensors and Bioelectronics 208):

114236.

doi:https://doi.org/10.1016/j.bios.2022.1 14236

- 45. Li C, Wang R, Wu A, Yuan T, Song K, Bai Y, Liu X (2022) SARS-COV-2 as potential microRNA sponge in COVID-19 patients. BMC Medical Genomics 15 (2): 1-10. doi:<u>https://doi.org/10.1186/s12920-022-01243-7</u>
- 46. Kara G, Arun B, Calin GA, Ozpolat B (2022) miRacle of microRNA-Driven Cancer Nanotherapeutics. Cancers 14 (15): 3818. doi:<u>https://doi.org/10.3390/cancers1415</u> <u>3818</u>
- 47. Meng F, Henson R, Lang M, Wehbe H, Maheshwari S, Mendell JT, Jiang J, Schmittgen TD, Patel T (2006) Involvement of Human Micro-RNA in Growth and Response to Chemotherapy in Human Cholangiocarcinoma Cell Lines. Gastroenterology 130 (7): 2113-2129. doi:https://doi.org/10.1053/j.gastro.2006. 02.057

 \bigcirc \bigcirc

Copyright © 2023 by the author(s). This is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<u>https://creativecommons.org/licenses/by/4.0/</u>).

How to Cite This Article:

Kanwal N, Al Samarrai OR, Al-Zaidi HMH, Mirzaei AR, Heidari MJ (2023) Comprehensive analysis of microRNA (miRNA) in cancer cells. Cellular, Molecular and Biomedical Reports 3 (2): 89-97. doi:10.55705/cmbr.2022.364591.1070

Download citation:

RIS; EndNote; Mendeley; BibTeX; APA; MLA; HARVARD; VANCOUVER